

# Immunomodulatory Effects of Sex Hormones: Requirements for Pregnancy and Relevance in Melanoma

Elizabeth Ann L. Enninga, BS; Sherman G. Holtan, MD;  
Douglas J. Crendon, MD, PhD; Roxana S. Dronca, MD; Wendy K. Nevala, MS;  
Simona Ognjanovic, PhD; and Svetomir N. Markovic, MD, PhD

## Abstract

Similarities between the pathologic progression of cancer and the physiologic process of placentation (eg, proliferation, invasion, and local/systemic tolerance) have been recognized for many years. Sex hormones such as human chorionic gonadotropin, estrogens, progesterone, and others contribute to induction of immunologic tolerance at the beginning of gestation. Sex hormones have been shown to play contributory roles in the growth of cancers such as breast cancer, prostate cancer, endometrial cancer, and ovarian cancer, but their involvement as putative mediators of the immunologic escape of cancer is still being elucidated. Herein, we compare the emerging mechanism by which sex hormones modulate systemic immunity in pregnancy and their potentially similar role in cancer. To do this, we conducted a PubMed search using combinations of the following keywords: “immune regulation,” “sex hormones,” “pregnancy,” “melanoma,” and “cancer.” We did not limit our search to specific publication dates. Mimicking the maternal immune response to pregnancy, especially in late gestation, might aid in design of better therapies to reconstitute endogenous antitumor immunity and improve survival.

© 2014 Mayo Foundation for Medical Education and Research ■ Mayo Clin Proc. 2014;89(4):520-535

## For editorial comment, see page 429

From Mayo Graduate School (E.A.L.E., S.O.), Department of Obstetrics and Gynecology (D.J.C.), Department of Oncology (R.S.D., W.K.N., S.N.M.), and Department of Medicine, Division of Hematology (S.N.M.), Mayo Clinic, Rochester, MN; and Oregon Health and Science University, Portland (S.G.H.).

In 1948, Beard and Krebs acknowledged a striking similarity between a trophoblast and a tumor, publishing their observation titled “The Unitarian or Trophoblastic Thesis of Cancer.”<sup>1-3</sup> Since then, these similarities have been extensively studied; many shared pathways and immunologic mediators have been identified.<sup>4,5</sup> The purpose of this review was to take an in-depth look at existing research describing the role of sex hormones in the potentially parallel settings of reproductive and tumor immunology, with a focus on metastatic melanoma. Although imperfectly understood, sex hormones are important regulators of the immune system in both pregnancy and cancer.<sup>6,7</sup> It is clear that they are involved in regulation and modification of the immune system to allow invasion, proliferation, and migration of tumor cells and trophoblasts.<sup>5</sup> It is possible that an organ system—level view of the process of placentation as well as melanoma progression could yield additional insights into potential therapeutic targets for hormone-based immune modulation.<sup>8</sup>

The complexities and redundancies involved in orchestration of the maternal response to pregnancy as well as the host response to cancer are increasingly appreciated.<sup>4,9</sup> Importantly, however, we and others have observed that neither pregnancy nor advanced cancers are static immunologic events.<sup>10-12</sup> Oscillations in systemic immunity between inflammation and tolerance seen in patients with metastatic melanoma have been documented and seem to follow a biologically predictable pattern. When tolerance seen in malignant melanoma is disrupted and brought back to a state of inflammation, patients have a much better prognosis than do those whose immune systems stay in an immunologically exhausted state. Pregnancy is also characterized by many hormonal fluctuations, although the time scale for these hormonal and immunologic changes may be measured in weeks as opposed to days<sup>13</sup> (Figure 1). Although it seems intuitive to consider the involvement of sex hormones interacting with the maternal immune system during pregnancy, it is less obvious but just as possible that such hormones alter systemic

immunity in the setting of cancers such as melanoma. We describe the observations and experimental evidence supporting such involvement in the following sections.

**CLINICAL EVIDENCE OF HORMONAL REGULATION OF MELANOMA**

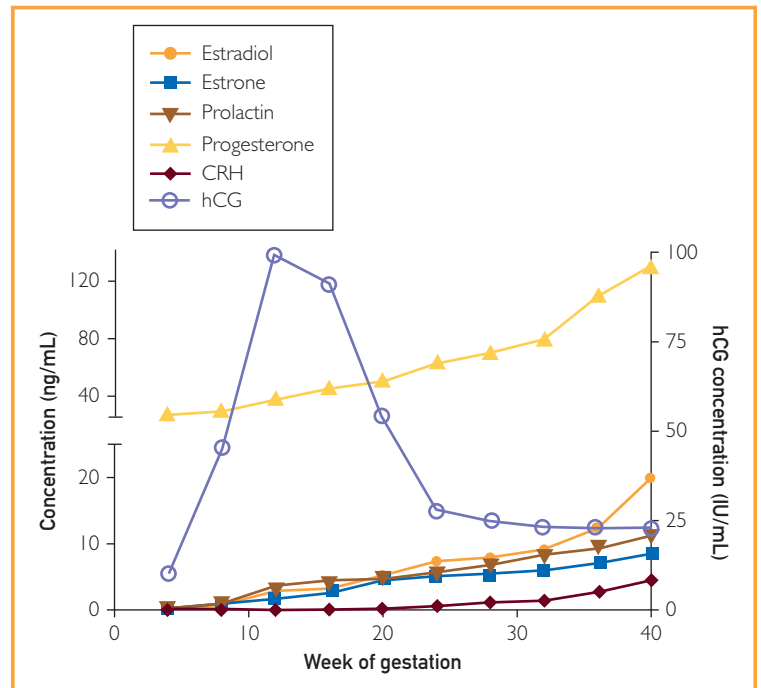
The skin is capable of producing many neuroendocrine mediators such as melanin, steroids, thyroid hormones, and sex hormones such as androgen, estrogen, and progesterin to maintain homeostasis; any failure to communicate between the skin, endocrine, and immune system could result in deregulation and disease.<sup>14,15</sup> Both melanocytes and melanoma tumors produce pigment in the melanosome that protects the skin against damaging ultraviolet rays through positive regulation by hormones such as L-tyrosine and L-dihydroxyphenylalanine.<sup>16,17</sup> Although the interplay between sex hormones and the immune system in melanoma remains poorly understood, several clinical observations support the role of sex hormones in melanoma development. Melanomas that are responsive to estrogens are associated with the superficial spreading melanoma subtype, a type of tumor with a much better prognosis. In addition, estrogen exerts a proliferative effect on melanocytes and can lead to the development of hyperpigmentation in women using oral contraceptives or hormonal replacement therapy.<sup>18</sup> Whether hormonal contraceptives increase the risk of melanoma is a matter of ongoing debate. Koomen et al<sup>19</sup> have reported that high levels of estrogens increase a woman's risk for developing malignant melanoma, while Lens and Bataille<sup>20</sup> have not observed a relevant association. It may be no coincidence that melanoma is the most common form of cancer associated with pregnancy.<sup>21</sup> This is believed to be due to the trophoblasts' increased need for lymphangiogenesis, which the melanoma then uses to promote its own growth. Complementary to this hypothesis, demographic characteristics and incidence may also provide an explanation as to this phenomenon. In addition, pregnant women are more likely than their nonpregnant counterparts to be diagnosed with an invasive melanoma.

We have shown that aging in healthy individuals is associated with a T<sub>H</sub>2 bias.<sup>22</sup> Women are more likely than men to develop melanoma before age 40, after which the diagnosis of

**ARTICLE HIGHLIGHTS**

- Systemic immunity in metastatic melanoma (cancer) mimics the systemic immune response of early pregnancy.
- Sex hormones promote/suppress different T cell responses during pregnancy and in melanoma.
- Metastatic melanoma and early pregnancy promote a systemic state of Th2 dominant chronic inflammation.
- Near parturition, hormones play a role in the return to cytotoxicity (Th1) of maternal immunity and the promotion of labor.
- Understanding the mechanism that causes immunity to switch from Th2 to Th1 in pregnancy may help researchers better understand how to break tolerance and improve patient outcomes in advanced cancers.

melanoma is observed at a much higher rate in men.<sup>23</sup> However, women diagnosed with melanoma have a better prognosis than do men, and premenopausal women have higher survival rates than do postmenopausal women.<sup>24,25</sup> Interestingly, melanoma metastasizes at a much slower rate in women than in men, and the pattern of metastatic spread is also different, with more



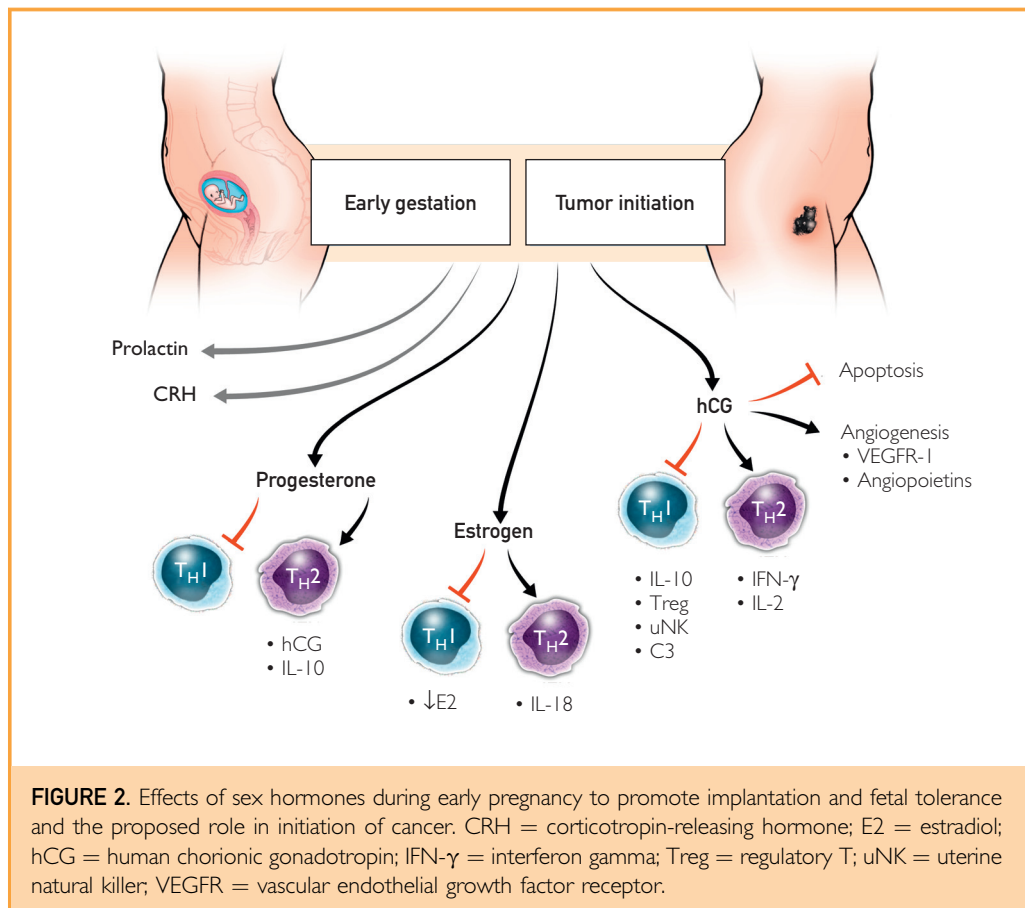
**FIGURE 1.** Hormonal changes (in weeks) important for the regulation of gestation in healthy pregnant women. CRH = corticotropin-releasing hormone; hCG = human chorionic gonadotropin.

locoregional recurrences observed in women.<sup>26,27</sup> Still, reasons behind these sex differences are yet to be elucidated. With an extensive amount of emerging data demonstrating the importance of sex hormones in immune function, in the remainder of the review we explore the mechanisms behind endocrine sex hormone-mediated immunomodulation. First, we highlight the potential involvement of hormones in tumor promotion, both via tolerance induction and as chronic inflammation and angiogenesis. Next, we describe hormones that may help improve antitumor immunity. Finally, we briefly discuss sexual dimorphism in immune responses, which may have implications for the development of personalized immunotherapy. A better understanding of immunologic switches that control tolerance, immune activation, and immune reconstitution, all of which can be studied using the different phases of pregnancy as a model, and could result in novel immunologic treatment strategies for melanoma and other malignancies.

## PUTATIVE PROTUMOR/IMMUNE-SUPPRESSIVE HORMONES: HUMAN CHRONIC GONADOTROPIN, PROGESTERONE, PLACENTAL GROWTH FACTOR, AND RELAXIN

### Human Chronic Gonadotropin

Soon after fertilization, the embryonic blastocyst begins secreting human chronic gonadotropin (hCG). Human chronic gonadotropin is a glycoprotein hormone primarily produced by trophoblasts that promotes many processes including implantation, recognition, differentiation, angiogenesis, and fetal-maternal homeostasis<sup>28</sup> (Figure 2). Levels of hCG continue to rise until the 11th week of gestation and then slowly decrease through the remainder of the pregnancy. The main purpose of hCG in pregnancy is to prevent degradation of the corpus luteum and stimulate progesterone production.<sup>29</sup> Human chronic gonadotropin also plays an important role in promoting immune suppression in the decidua by preventing



**FIGURE 2.** Effects of sex hormones during early pregnancy to promote implantation and fetal tolerance and the proposed role in initiation of cancer. CRH = corticotropin-releasing hormone; E2 = estradiol; hCG = human chorionic gonadotropin; IFN- $\gamma$  = interferon gamma; Treg = regulatory T; uNK = uterine natural killer; VEGFR = vascular endothelial growth factor receptor.

maternal macrophage phagocytosis to the invading trophoblast to establish immune tolerance.<sup>30</sup> The early trophoblast promotes fetal tolerance by secreting hCG, which acts as a powerful chemoattractant for regulatory T (Treg) cells to migrate to the placenta after fertilization.<sup>31</sup> Treg cells play an important role in maintaining self-tolerance and modulating tolerance to nonself antigen, such as those displayed by the fetus. Expression of complement component 3 was found to be up-regulated by hCG on stromal cells of the baboon endometrium postovulation, suggesting that hCG is also able to modulate the decidual environment during the preimplantation stage.<sup>32,33</sup>

Human chronic gonadotropin also regulates uterine natural killer (uNK) cells.<sup>34</sup> This uNK cell subset makes up approximately 70% of the lymphocyte population in the endometrium and plays an important role in maintaining and regulating the uterine spiral arteries during the first trimester.<sup>35</sup> Uterine natural killer cells stimulate decidual monocytes to secrete interferon gamma (IFN- $\gamma$ ), promoting Treg-cell proliferation through indoleamine 2,3-dioxygenase and transforming growth factor beta.<sup>36</sup> Interestingly, we have found uNK cells in the blood of patients with stage IV melanoma and they also positively correlated with transforming growth factor beta levels in plasma.<sup>37</sup> Human chronic gonadotropin drives a systemic response during pregnancy. Before in vitro fertilization, women given hCG had increased levels of anti-inflammatory IL-27 and IL-10 and reduced levels of pro-inflammatory IL-17, which resulted in an increase in the number of Treg cells and a more receptive uterine wall for implantation.<sup>38</sup> They also notice that hCG affected the maternal adaptive immune system, promoting a T<sub>H</sub>2-differentiated state by activating T cells that produce IL-4 while inhibiting T cells that secrete IFN- $\gamma$ .<sup>38</sup>

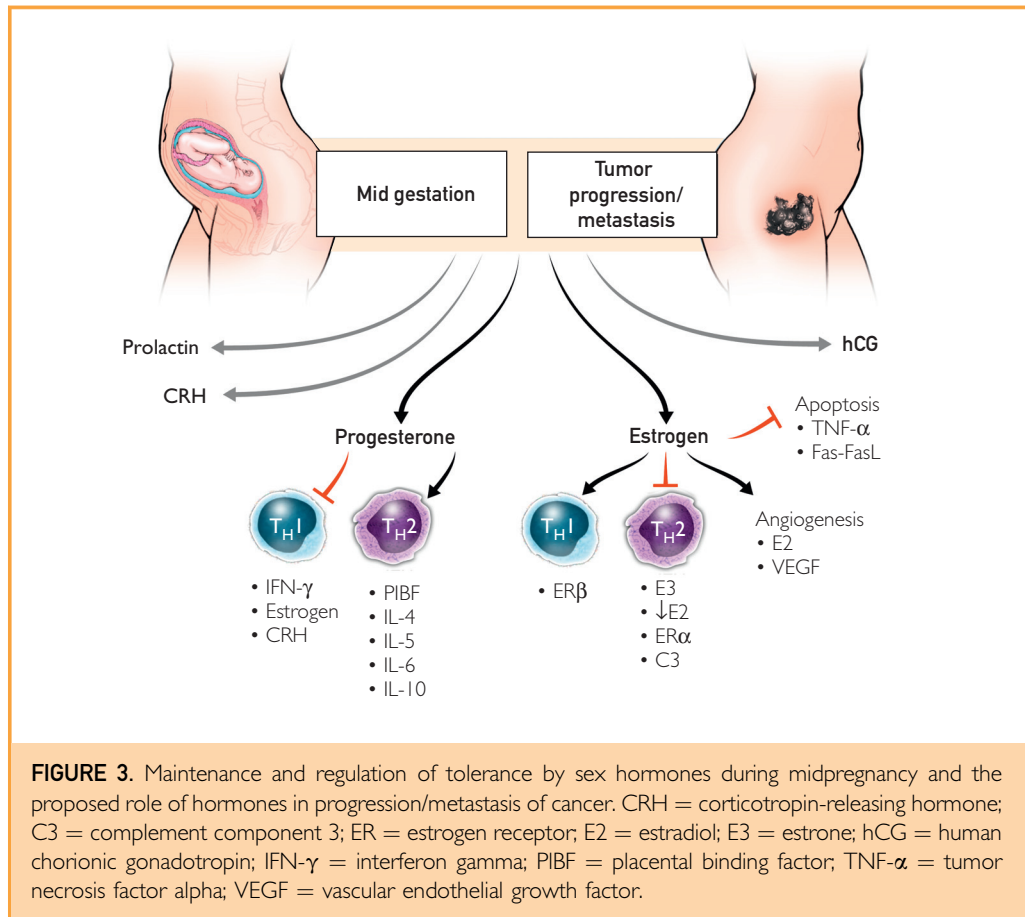
In addition, the formation of new blood vessels is also driven by hCG, which acts on proangiogenic molecules such as vascular endothelial growth factor (VEGF) receptor 1 and angiopoietins.<sup>39</sup> Soluble VEGF-C aids in immune tolerance by suppressing the cytotoxic activity of uNK cells at the fetal-maternal interface.<sup>40</sup> Human chronic gonadotropin enhances VEGF production by endometrial cells through paracrine feedback, further promoting blood vessel formation to the fetus.<sup>41</sup>

IL-10, an inhibitor of inflammatory cytokines that is found at high levels near the beginning of pregnancy, induces the trophoblast to produce VEGF-C and stimulate placental angiogenesis.<sup>42</sup> Also, endocrine gland-derived VEGF has been implicated as a negative regulator of trophoblast invasion in the placenta, as high levels of endocrine gland-derived VEGF are seen in preeclampsia, and has recently been hypothesized to be regulated by hCG.<sup>43</sup> Human chronic gonadotropin prevents apoptosis of endometrial cells through the Fas-FasL pathway, which, in turn, drives maternal tolerance during early pregnancy.<sup>44</sup>

Human chronic gonadotropin plays very similar roles in the invasion and progression of cancer (melanoma). The hCG receptor was first discovered in trophoblastic neoplasms, which suggested that it has an important role in regulating growth and invasion not only in pregnancy but in cancer as well.<sup>45</sup> Several studies have found that many cancers, including bladder cancer, cervical cancer, lung cancer, pancreatic cancer, and colorectal cancer, can be diagnosed by high levels of hCG in the serum.<sup>46,47</sup> Human chronic gonadotropin mRNA has also been expressed in tumor cells from patients diagnosed with malignant melanoma and has been suggested for use as a biomarker for disease.<sup>48</sup> Others have shown that up to 60% of active neoplasia will express high levels of hCG in serum.<sup>49</sup> Antibodies against hCG have been detected during malignancy, but hCG levels are so high that the effect of these antibodies seems to be minimal, thereby allowing hCG to act as an autocrine growth factor to promote malignancy.<sup>50</sup> Tumor secretion of hCG prevents apoptosis, allowing the cancer to become more resistant and aggressive.<sup>51</sup> As a potent angiogenic factor, hCG secreted by the tumor stimulates sprout formation through vasodilation, maturation, and increased vessel permeability, thus promoting tumor growth.<sup>52</sup>

### Progesterone

Initial production of progesterone in pregnancy is induced by hCG. Progesterone secretion by the placenta continues to increase throughout pregnancy, only slightly decreasing approximately 4 weeks before labor onset. Progesterone is a potent immunomodulator that establishes T<sub>H</sub>2 bias in pregnancy by reducing the production of pro-inflammatory cytokines by macrophages in



response to infectious stimuli, and altering cytokine secretion of T-cell subsets toward IL-10 production<sup>53</sup> (Figures 2 and 3). Furthermore, it induces secretion of chemokines such as CXCL10, CX3CL1, and CCL2 that localize T<sub>H</sub>2-biased immune cells to the placenta and up-regulates nonclassical human leukocyte antigen-G.<sup>54</sup> Interestingly, most progesterone receptor (PR)-expressing decidual T cells express a  $\gamma/\delta$  T-cell receptor.<sup>55</sup> This limits the number of ligands the T-cell receptor can recognize, which provides protection to the growing fetus. These T cells are also believed to be able to identify antigens presented by trophoblasts.<sup>56</sup> The PR comes in 2 main isoforms, PR-A and PR-B, which compete with one another for progesterone binding.<sup>57</sup> Progesterone receptor-B is expressed on myometrial cells during most of pregnancy and inhibits the expression of pro-inflammatory genes; however, PR-A is expressed during labor and it in turn promotes pro-inflammatory gene activation.<sup>58</sup> Progesterone receptor-A overexpression at the end of pregnancy also promotes the

activation of estrogen by increasing the expression of estrogen receptor (ER)  $\alpha$ .<sup>59</sup> A smaller, soluble PR isoform, PR-C, has been found to compete specifically with PR-B for progesterone binding, by binding directly to progesterone and inhibiting PR-B signaling near parturition, thereby promoting labor-associated myometrium changes and activating pro-inflammatory pathways.<sup>60</sup>

The immune effects of progesterone are exerted by inhibition of pro-inflammatory transcription factor nuclear factor kappa B (NF- $\kappa$ B) through I $\kappa$ B kinase.<sup>61</sup> Effects of progesterone are also mediated by progesterone-induced blocking factor, which is expressed on lymphocytes in the decidua and is important in maintaining pregnancy. Women who suffer from spontaneous miscarriages or have high stress levels have been found to have low levels of progesterone-induced blocking factor, which was associated with pregnancy complications.<sup>62</sup> Natural killer (NK) cells in pregnancy are very sensitive to progesterone: low levels of

progesterone are required to achieve inhibition, compared with 100-fold higher levels needed to achieve similar NK-cell inhibition in non-pregnant individuals.<sup>63</sup> In mice, progesterone activates a T<sub>H</sub>2-biased immune response by inhibiting maturation of dendritic cells (DCs), which would more readily initiate a T<sub>H</sub>1 response through the promotion of cytotoxic T-cell expansion.<sup>64</sup>

In cancer, progesterone modulates immune responses by inhibiting T-cell proliferation<sup>65</sup> and IFN- $\gamma$  expression<sup>66</sup> while enhancing IL-4, IL-5, IL-6, and IL-10 production and promoting a humoral response.<sup>67</sup> Interestingly, human melanocytes can produce steroids *de novo* through the metabolism of progesterone or cholesterol.<sup>68-70</sup> Progesterone has been shown to inhibit the proliferation of human melanocytes by blocking the effects of estrogen.<sup>71</sup> The PR has been identified in the cytoplasm and nucleus of melanocytes by immunohistochemistry.<sup>72</sup> Melanoma cells that do not express the PR were still found to be regulated in the presence of progesterone, but are modulated through signal transduction versus transcription.<sup>73</sup> Progesterone-induced blocking factor has been suggested to play a role in cell cycle regulation and T<sub>H</sub>2-biased immunity through the IL-4 receptor.<sup>74</sup> Progesterone-induced blocking factor mRNA is constitutively expressed in tumor cells and does not require the presence of the PR, which provides a mechanism for cancer to escape antitumor immune responses.<sup>56</sup> *In vitro* experiments using WM266-4 cells found that progesterone, estradiol 17 $\beta$  and dihydrotestosterone given together can inhibit tumor growth through the down-regulation of IL-8, which is a potent cytokine for inducing melanoma cell growth.<sup>75</sup>

### Placental Growth Factor

We have previously described a VEGF-driven state of chronic inflammation in metastatic melanoma.<sup>76</sup> Placental growth factor (PlGF), a VEGF homologue, may play a similar role in perpetuating chronic inflammation. It plays a considerable role in embryogenesis, promoting both angiogenesis and vascularization to the fetus during inflammation.<sup>77</sup> It was discovered that Flt-1, a VEGF receptor, binds PlGF and mediates recruitment of monocytes.<sup>78</sup> Peripheral blood monocytes treated with PlGF showed increased expression of pro-inflammatory cytokines IL-1 $\beta$  and tumor necrosis factor alpha (TNF- $\alpha$ ),

and chemokines monocyte chemoattractant protein, IL-8, and macrophage inflammatory protein, suggesting that PlGF plays an important role in inducing inflammation.<sup>79</sup> Melanocytes and melanoma tumors are also known to secrete PlGF, which makes them weakly responsive to anti-VEGF therapy.<sup>80</sup> However, when PlGF was neutralized, even in tumors resistant to anti-VEGF therapy, the tumor could no longer signal through vascular endothelial growth factor receptor-1, inhibiting growth during a preclinical study.<sup>81</sup> Therefore, it has been hypothesized that PlGF plays an important role in allowing a tumor to become drug resistant.<sup>82,83</sup> Further studies need to be conducted to fully understand PlGF's role in inflammation and tumor progression.

### Relaxin

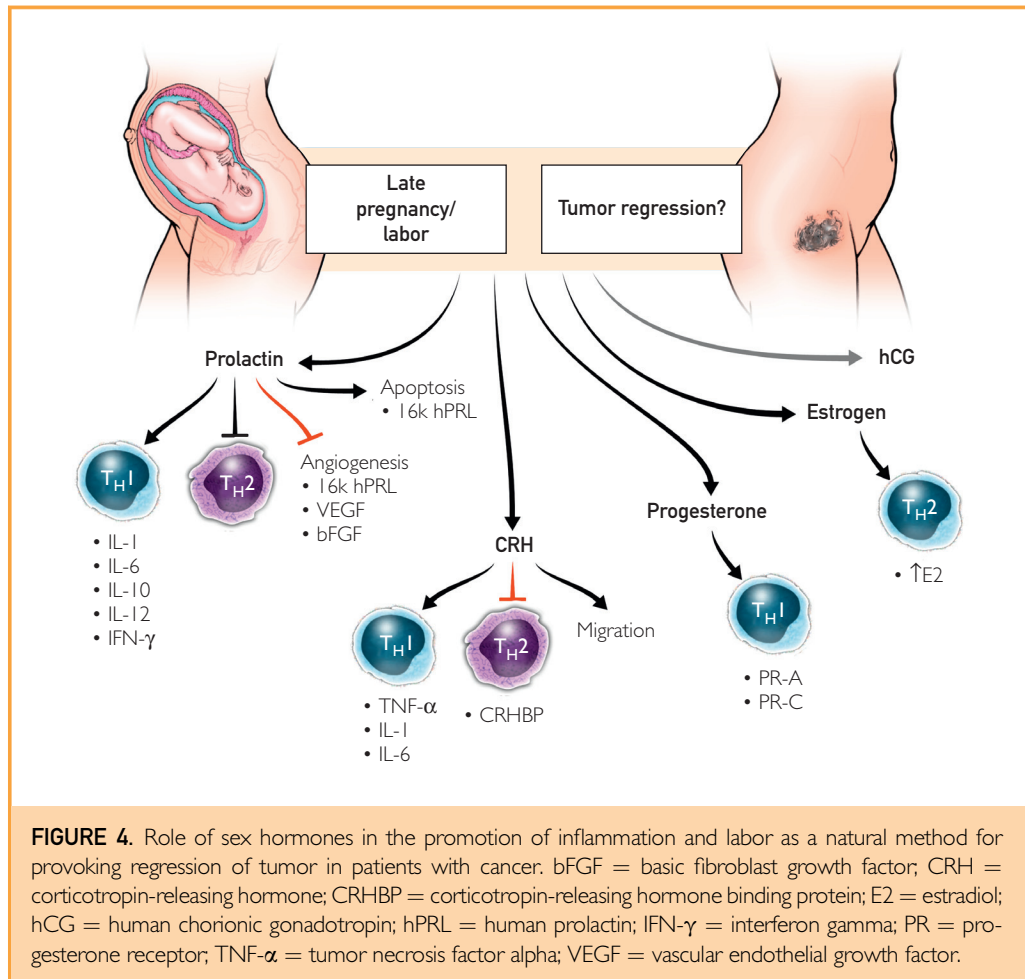
The role of relaxin in pregnancy (induction of matrix metalloproteinases, extracellular matrix remodeling, labor), as well as induction of pro-inflammatory cytokines IL-6 and IL-8, has been well documented in rhesus monkeys.<sup>84</sup> Although relaxin has not been studied in melanoma to date, its role in carcinogenesis of breast and prostate cancer has been well established. Relaxin was implicated in tumor growth, cell invasion during pregnancy, and likely tumor invasion in carcinogenesis as well as in angiogenesis by the induction of VEGF.<sup>85-87</sup> Whether relaxin plays a role in melanoma development remains to be elucidated.

### PUTATIVE ANTITUMOR/PRO-INFLAMMATORY HORMONES: CORTICOTROPIN-RELEASING HORMONE, PROLACTIN, AND VISFATIN

#### Corticotropin-Releasing Hormone

Corticotropin-releasing hormone (CRH) has been postulated to regulate the duration of gestation, with its levels being the highest during labor (Figure 4). It works directly on myometrial cells to facilitate the onset of labor, and it is tightly regulated by progesterone, estrogens, nitric oxide, IL-1 $\beta$ , and TNF- $\alpha$ .<sup>60</sup> Corticotropin-releasing hormone binding protein binds CRH and inactivates its ability to promote corticotropin production.<sup>88</sup> Corticotropin-releasing hormone binding protein levels decrease throughout pregnancy, allowing high levels of CRH to accumulate and promote labor. Other actions of CRH include regulation of





fetal blood flow, placental prostaglandin and cortisol production, and uterine contractility.<sup>89</sup>

Corticotropin-releasing hormone regulates the hypothalamic-pituitary-adrenal (HPA) axis, which allows cells to respond to environmental stresses.<sup>90</sup> Melanocytes are known to secrete CRH, which permits them to produce cortisol and corticotropin to maintain skin homeostasis.<sup>91</sup> In cell culture, melanoma cells treated with CRH migrated further during stress, which was determined to be mediated by the extracellular signal-regulated kinase (ERK1/2) pathway.<sup>92</sup> Increases in the levels of CRH and proopiomelanocortin are strongly associated with malignant melanoma.<sup>93</sup> Another function of CRH in the skin is to act as a growth regulator by both promoting and suppressing cell proliferation. Corticotropin-releasing hormone receptor 1 controls the action of CRH and can promote cyclic adenosine monophosphate and

inositol trisphosphate synthesis in dermal and epidermal cells.<sup>94</sup> Melanomas exclusively express corticotropin-releasing hormone receptor 1, which plays an important role in proliferation and has become an agonist target.<sup>95</sup> In a B16 mouse melanoma model, daily injections with CRH reduced tumor volume by 30% to 60% compared with that in control animals.<sup>96</sup> Research shows that CRH promotes the survival of melanocytes during starvation and prevents cell proliferation by inhibiting growth factor signaling through cyclic adenosine monophosphate and IP3 second messengers.<sup>97</sup> Tumor necrosis factor alpha, IL-1, and IL-6 stimulate the HPA axis to produce CRH, which drives a pro-inflammatory, T<sub>H</sub>1-biased response.<sup>98</sup> When peripheral CRH acts on the HPA axis in the brain, it triggers a classical feedback mechanism that leads to the secretion of cortisol, a steroid hormone with anti-inflammatory

effects.<sup>99</sup> Thus, CRH is a potential modulator of a T<sub>H</sub>1-biased response and its role should be further studied in cancer.

### Prolactin

Prolactin (PRL) is a polypeptide hormone secreted by the syncytiotrophoblast that reaches its highest levels during late pregnancy (Figure 4). It acts on corpus luteum cells in the ovary to stimulate the secretion of progesterone to maintain pregnancy and on epithelial cells of the mammary gland to initiate milk production.<sup>100</sup> The amount of PRL secreted is directly proportional to the size of the fetus, and it functions to provide energy for the mother and nutrients for the fetus. Through lipolysis and anti-insulin effects of PRL action, the maternal insulin level increases, providing free fatty acids and amino acids to the growing fetus.<sup>101</sup> Prolactin functions to increase  $\beta$ -islet cell proliferation, inhibit apoptosis, and cause  $\beta$ -islet cells to become more responsive to glucose.<sup>102</sup> Glucocorticoid expression of Rasd1 near the end of gestation changes insulin secretion during pregnancy through the inhibitory effects of PRL on Rasd1 transcription.<sup>103</sup> Activation of dopamine neurons suppresses PRL secretion through placental lactogens.<sup>104</sup> The N-terminal 16K PRL fragment had been determined to stop proliferation and migration of vascular endothelial cells and cause cell cycle arrest resulting in apoptosis.<sup>105</sup> High levels of 16K human PRL can be detected in serum and urine of women suffering from preeclampsia.<sup>106</sup>

In melanoma, PRL drives T<sub>H</sub>1-biased immunity through the secretion of IL-1, IL-6, IL-10, IL-12, and IFN- $\gamma$  by NK cells and B lymphocytes,<sup>107,108</sup> and plasma cell activation.<sup>109</sup> It increases T-cell proliferation<sup>110</sup> and decreases B-cell apoptosis.<sup>111</sup> Through animal studies, PRL's antitumor effect has been shown to promote tumor-specific macrophages through IFN- $\gamma$  and IL-12, and CT26 tumor-bearing mice injected with recombinant human PRL and IL-15 had enhanced cytotoxic activity to the tumor, resulting in fewer lung metastasis and longer overall survival (OS).<sup>112,113</sup> However, PRL does not have an antitumor effect in all cancers. In breast cancer, high levels of PRL are associated with a greater risk for developing cancerous ER $\alpha$ + tumors and is associated with much poorer outcomes for patients with these increased levels.<sup>114</sup> This is likely because PRL

promotes mammary tumorigenesis independent of cyclin D1 activation.<sup>115</sup> This has not yet been observed in melanoma. Pro-inflammatory cytokines such as IL-1, IL-2, and IL-6 produced by both pituitary and extrapituitary cells have a stimulatory effect on PRL secretion, while IFN- $\gamma$  inhibits its production.<sup>116</sup> Many immune cells express the PRL receptor including monocytes, macrophages, B and T cells, granulocytes, and NK cells.<sup>117</sup> Prolactin secretion from the anterior pituitary gland is inhibited by the dopamine D2 receptor, which is widely expressed on melanoma cells and plays a key role in inhibiting adenylyl cyclase, which is necessary for cellular signal transduction.<sup>118,119</sup> Prolactin also enhances the effect of immune cells, including CD34<sup>+</sup> stem cells, in the blood through the up-regulation of major histocompatibility complex class II expression on antigen-presenting cells, T-cell clonal expansion, antibody production, increased cytotoxicity of NK cells, and microbe killing by macrophages.<sup>120</sup> In mouse melanoma models, 16K human PRL can block angiogenesis and inhibit tumor growth by activating NF- $\kappa$ B, causing tumor-infiltrating lymphocytes to access and destroy cancer cells.<sup>121</sup> This PRL peptide endogenously blocks Notch signaling, which greatly impairs the tumor's ability to vascularize.<sup>122</sup> 16K human PRL has also been shown to block angiogenesis through the inhibition of basic fibroblast growth factor and VEGF, making it a target for antitumor therapies.<sup>123</sup> Thus, better understanding of how PRL can promote cell-mediated immunity could help researchers design better ways to initiate tumor destruction.

### Visfatin

Previously known as pre-B-cell colony-enhancing factor or nicotinamide phosphoribosyltransferase, visfatin is a visceral fat cytokine with an important role in the promotion of inflammation.<sup>124-126</sup> Visfatin levels are highest at the end of gestation and promote the secretion of IL-1 $\beta$ , IL-6, IL-8, cyclooxygenase 2, TNF- $\alpha$ , and prostaglandin E<sub>2</sub>.<sup>127,128</sup> Interestingly, infection-associated preterm labor is concomitant with elevated levels of visfatin in maternal plasma.<sup>129</sup> These findings suggest that visfatin could be important for immune resolution back to a T<sub>H</sub>1 state. However, the role of visfatin in melanoma is poorly understood. It appears that visfatin is more highly expressed in melanoma lesions than in benign



lesions.<sup>130</sup> Moreover, it seems that visfatin may promote melanoma cell growth in vitro. Namely, a study<sup>131</sup> using the melanoma Me45 cell line reported that visfatin increased the proliferation of these tumor cells. In another study,<sup>132</sup> neutrophils have been found to synthesize visfatin in response to inflammatory stimulus and inhibit apoptosis in vitro and in patients with sepsis. Thus, it remains possible that there could be an additional regulatory level of visfatin during pregnancy, which might be lost during tumorigenesis, and thereby promotes melanoma proliferation, but more studies need to be conducted to elucidate the role of visfatin in melanoma.

### ESTROGEN: A DOUBLE-EDGED SWORD MODULATING T<sub>H</sub>1/T<sub>H</sub>2 IMMUNITY

Women have high levels of estrogen, which drop dramatically when they reach menopause. Estrogen modulates the immune response by inducing peripheral T cells to secrete pro-inflammatory cytokines IFN- $\gamma$  and IL-2,<sup>133</sup> but also promotes tolerance by inducing IL-10 secretion<sup>134</sup> (Figures 3 and 4). Estradiol-17 $\beta$  (E2) at high concentrations induces a T<sub>H</sub>1 response, whereas at low concentrations it biases the system toward immune tolerance.<sup>135</sup> Pro-inflammatory responses are regulated by E2 through NF- $\kappa$ B. However, estrone (E3) is detectable only in pregnant women because it is produced by the placenta and fetus. Estrone is important for reducing the production of pro-inflammatory cytokines in circulation by decreasing I $\kappa$ B degradation, which inhibits NF- $\kappa$ B activation and apoptosis (Figure 3).<sup>136</sup> Estrogens work through interaction with the ER to induce transcriptional regulation. Immune cells including DCs, NK cells, macrophages, and lymphocytes express ERs, signifying that this hormone modulates their function.<sup>137</sup> Indeed, the differentiation of DCs is regulated by E2 acting on ER $\alpha$ . Levels of E2 are the highest during the third trimester, when the number of immature DCs is high and actively presenting fetal antigen to the mother's immune system to begin labor.<sup>138</sup> Complement component 3, a protein that plays a central role in inducing inflammation in innate immunity, is increased by estrogen in oviductal epithelial cells.<sup>139</sup> Angiogenesis in the uterus is also driven by estrogen. Vascular endothelial growth factor mRNA levels increase in the endometrium in the presence of E2.<sup>140</sup> This

increases placental blood flow and vasodilation, which are both characteristic of angiogenesis. Cytotrophoblasts, but not syncytiotrophoblasts, stimulate VEGF production directly, and this production is correlated to the levels of E2 in the serum.<sup>141</sup> Thus, estrogen stimulates VEGF production and blood vessel formation, which is essential for the establishment and maintenance of the fetus during pregnancy.

Women at risk for familial breast cancer have increased risk for developing melanoma and vice versa, suggesting that estrogens can promote tumorigenesis.<sup>142</sup> It has been known for many years that melanoma tumor cells express ERs.<sup>143</sup> Estrogens affect lymphocytes by initiating the secretion of IL-10, IL-12, and IFN- $\gamma$  and inhibition of TNF- $\alpha$ ,<sup>144</sup> stimulation of antibody production,<sup>145</sup> and the reduction of macrophage and DC apoptosis.<sup>146,147</sup> Estradiol-17 $\beta$  has been proven to inhibit melanoma growth by obstructing receptor binding of IL-8.<sup>75</sup> Chronic exposure to estrogens was suggested to increase NF- $\kappa$ B stimulation and induce pro-inflammatory responses by macrophages.<sup>148</sup> It has been shown that short-term and long-term inflammation leads to the up-regulation of ER $\alpha$ , but not ER $\beta$ , thus affecting the effects of estrogens on T cells, because T cells express ER $\alpha$ , while B cells express ER $\beta$ .<sup>149</sup> It was also reported that estrogen decreases apoptosis, as well as TNF- $\alpha$  production.<sup>150</sup> Estradiol-17 $\beta$  and estrone both have a strong affinity for binding ER $\alpha$ ,<sup>151</sup> which is why ER $\alpha$  is commonly associated with tumor promotion. Yet, ER $\beta$  has been found on malignant melanoma cells that are negative for ER $\alpha$ .<sup>152</sup> Loss of ER $\beta$  expression correlated with increased invasiveness of the tumor,<sup>153</sup> suggesting that loss of ER $\beta$  expression increases malignant transformation in melanoma. The most important prognostic factors in melanoma are tumor thickness (Breslow depth) and invasive level (Clark level); as each increase, the patient's prognosis decreases.<sup>154</sup> Interestingly, increased ER $\beta$  expression is not found on nonmalignant cells surrounding the tumor, only on the tumor cells themselves.<sup>155</sup> Another role of ER $\beta$  is regulation of monocyte apoptosis through the Fas-FasL signaling pathway.<sup>156</sup> It has been shown that myeloid progenitor cells exposed to the granulocyte-macrophage colony-stimulating factor differentiate into immature DCs through the expression of ER $\alpha$  and increased levels of estrogens.<sup>157,158</sup>

Recently, 2-methoxyestradiol, an estrogen derivative, has been determined to be a potent inhibitor of angiogenesis and melanoma growth in a mouse model.<sup>159</sup> Taken together, this indicates that estrogens can both promote and hinder tumor growth and monitoring ER expression could help clinicians determine the patient prognosis to the disease.

### SEXUAL DIMORPHISM IN RESPONSE TO HORMONES

In this review, we have discussed existing data supporting a role for sex hormones and their ability to manipulate the immune system in both pregnancy and melanoma. Beyond these observations, several studies have identified sexual dimorphism with respect to inflammatory responses in various settings. For example, sex-based differences in vascular function have been described and show the development of early atherosclerotic lesions and plaques with increased production of inflammatory mediators (IL-10 and TNF- $\alpha$ ) in women than in men.<sup>160-162</sup> That the state of pregnancy, and not a general state of tolerance in women, accounts for additional sex-based differences in immune function is supported by the fact that autoimmune diseases are more prevalent in women.<sup>163</sup> Moreover, autoimmune diseases such as rheumatoid arthritis and systemic lupus erythematosus are diagnosed more frequently in women and are strongly correlated with sex hormones.<sup>164,165</sup> Interestingly, some autoimmune diseases such as arthritis remit with pregnancy, but systemic lupus erythematosus has been found to often worsen in severity during gestation.<sup>164</sup> Women also exhibit enhanced innate immune responses to infections,<sup>166,167</sup> not blunted ones, and survive episodes of severe sepsis to a greater degree than do men.<sup>168</sup> It is clear that sex plays a role in risk, severity, and prognosis of many diseases, including cancer, and research in this area has just begun. It may one day become important to consider the sex of the patient with cancer when personalizing cancer immunotherapies.

### REVERSING TOLERANCE IN MELANOMA

To positively affect survival of patients with cancer, it appears critical to find a way to break immune tolerance to the malignancy. Insights

into achieving this goal in metastatic melanoma may lie in well-recognized abnormalities of pregnancy (eg, spontaneous abortions). For example, serum taken from women suffering from recurrent spontaneous abortion have a cytokine profile indicative of a T<sub>H</sub>1 response compared with healthy pregnant women.<sup>169</sup> To counter this, intravenous immunoglobulin given to women, with reported previous recurrent miscarriages and/or suffering from recurrent miscarriages after in vitro fertilization, every 3 to 4 weeks starting from conception or 24 hours before embryo transfer showed a decrease in the number of NK cells and an implantation success rate of 92.5% compared with 25% for women not receiving intravenous immunoglobulin.<sup>170</sup> Moreover, new immunotherapies for promoting tolerance to the fetus during gestation are being studied in animal models and could have important implications in melanoma. In mice, researchers have shown that blocking CD80 and CD86 enhances maternal tolerance, decreasing implantation failure through the increase in Treg cells and development of T<sub>H</sub>2 cells.<sup>171</sup> The use of anti-cytotoxic T lymphocyte antigen 4 (CTLA4) antibody therapy with CD80/CD86 blockade has been found to regulate the T<sub>H</sub>2/T<sub>H</sub>1 balance in peripheral blood monocytes isolated from women with recurrent spontaneous abortion, which led to the design of an adenoviral CTLA4 antibody that improved pregnancy outcomes in a mouse model of spontaneous abortion.<sup>172,173</sup>

The opposite, immune-activating effect is desired in metastatic melanoma. Recent Food and Drug Administration approval of the anti-CTLA4 antibody, ipilimumab, was meant to initiate T-cell activation and tumor destruction by promoting cytotoxic capacity of naturally occurring tumor-specific T cells, thereby overcoming, in part, their state of immune tolerance.<sup>174</sup> Patients with metastatic melanoma treated with ipilimumab given alone or in combination with a peptide vaccine (gp100) exhibited a median overall survival OS time of approximately 10 months, an improvement over the 6.4-month OS observed in patients receiving gp100 vaccine alone.<sup>175</sup> As breakthrough as this drug is, it comes at a cost of significant immune-related adverse events in up to 15% of the patients.<sup>176</sup> Another new and promising antibody targets program death 1 (PD-1), an additional receptor

found on activated cells that is critical for immune regulation, which has been shown to promote a pro-inflammatory environment by IFN- $\gamma$  and IL-2 secretion.<sup>177</sup> In clinical trials, this drug, after treatment with or without the anti-CTLA inhibitor, showed a high rate of sustained tumor regression and a median OS of 11 months in patients with metastatic melanoma.<sup>178</sup> This led to administering both ipilimumab and nivolumab (anti-PD-1) concurrently, resulting in 53% of the patients obtaining an 80% reduction in tumor volume.<sup>179</sup> Unfortunately, the reported increase in objective response rate was paralleled with a similar increase in severe toxicity. Other promising immunotherapies for patients with melanoma are also being studied. Antitumor immunity has been shown by using an antibody to CCR4, a marker found on immune-suppressive Treg cells, which when given to a patient with T-cell leukemia-lymphoma resulted in a CD8<sup>+</sup> T-cell response to the tumor.<sup>180</sup> Human epidermal growth factor receptor 2, an antibody commonly used to treat breast cancers, has recently shown positive effects on a number of melanoma cell lines and xenograft models.<sup>181</sup> Bevacizumab, an anti-VEGF antibody, given to patients with metastatic melanoma in combination with albumin-bound paclitaxel and carboplatin, resulted in an increase in CD8<sup>+</sup> lymphocytes but did not affect the T<sub>H</sub>1/T<sub>H</sub>2 ratio.<sup>182</sup>

Even though promising, the clinical successes of present day immunotherapeutic strategies for metastatic cancer fall short of their preclinical results. Many share the belief that the reason for this discrepancy in clinical translation is the result of tumor-driven immune tolerance of human cancer.<sup>183</sup> Another reason for this discrepancy is that malignant melanocytes use melanogenesis, which is a normal metabolic process that generates a local immunosuppressive environment through proopiomelanocortin-derived peptides and steroids.<sup>17</sup> It was discovered that inhibiting melanogenesis increases the potency of the immune system and chemotherapy against tumors.<sup>184</sup> Overcoming the tumor-induced modulation of systemic immunity to recover the ability of endogenously generated immune cells to effectively destroy the malignancy will be a considerable challenge. An interesting concept in pregnancy, which could lead to better understanding of malignancy, is the

spontaneous return to cytotoxicity near the end of parturition.<sup>138</sup> Near parturition, an unknown event results in the reactivation of T<sub>H</sub>1 maternal immunity and the initiation of labor (rejection). Characterizing this phenomenon by further studying the role that cytokines, cells, and hormones play could translate into different methods for breaking tolerance in patients with cancer, which could ultimately improve therapeutic efficacy and OS. The skin is a steroidogenic organ; it synthesizes its own steroids and sex hormones and can regulate local immune activities along with affecting the function of the epidermis.<sup>185</sup> Corticotrophin-releasing hormone has been found to block human melanoma cell proliferation *in vitro*<sup>97</sup> and could provide additional therapeutic value when paired with a targeted agent. As progesterone promotes a tolerant state in pregnancy, treating a stable melanoma patient with progesterone was found to cause proliferation of dormant micrometastases by tipping the immune system back to a T<sub>H</sub>2 state.<sup>186</sup> B7-H1 (PD-L1) is a molecule expressed on antigen-presenting cells and contributes to tumor evasion and expansion of Treg cells.<sup>187,188</sup> In a B7-H1 knockout mouse model of melanoma, it was discovered that females had superior tumor growth resistance than did males due to estrogen-mediated suppression of Treg cells.<sup>189</sup> Clinically, in a phase I study of advanced cancers, including melanoma, patients treated with the PD-1 antibody reported a marked response rate (28%) that was durable in only those patients expressing PD-L1, making it a potential biomarker for anti-PD-1 treatment response.<sup>190</sup> However, the authors did not compare men and women to look at potential differences due to sex. Altogether, to improve outcomes for patients with melanoma treated with immunotherapeutics, we must more completely understand the process of normal pregnancy immunoregulation, specifically the return to cytotoxicity (T<sub>H</sub>1 bias) at the end of gestation.

## CONCLUSION

The dynamic maternal immune responses to normal pregnancy have evolved out of the need to support a semiallogenic fetus over the duration of the pregnancy, without relevant infectious or inflammatory impediment to the mother. In turn, the maternal immune system

is tightly regulated by hormone release and cytokine action to protect the developing fetus. Cancers, including melanoma, appear to induce similar tolerogenic immune programs through various mechanisms, including paracrine secretion of sex hormones, to drive angiogenesis required for oxygen and nutrient supply, all the while evading immune attack in a manner similar to the process of placentation. A better understanding of the molecular switches involved in the induction and reversal of immune tolerance in the setting of pregnancy may help identify new methods for targeted immune modulation for patients with melanoma.

**Abbreviations and Acronyms:** CRH = corticotropin-releasing hormone; CTLA4 = cytotoxic T lymphocyte antigen 4; IFN- $\gamma$  = interferon gamma; DC = dendritic cell; E2 = estradiol-17 $\beta$ ; ER = estrogen receptor; hCG = human chorionic gonadotropin; HPA = hypothalamic-pituitary-adrenal; NF- $\kappa$ B = nuclear factor kappa B; NK = natural killer; OS = overall survival; PD-1 = program death 1; PlGF = placental growth factor; PR = progesterone receptor; PRL = prolactin; TNF- $\alpha$  = tumor necrosis factor alpha; Treg = regulatory T; uNK = uterine natural killer; VEGF = vascular endothelial growth factor

**Grant Support:** This project was supported by the Office of Research in Women's Health and grant BIRCWH Award Number 2K12HD043488-11 (S.G.H.) from the National Institute of Child Health and Human Development, Oregon and NIH/NCATS CTSA grant number TLI TR000137 (E.A.L.E.).

**Correspondence:** Address to Svetomir N. Markovic, MD, PhD, Division of Hematology, Department of Medicine, Mayo Clinic, 200 First St SW, Rochester, MN 55905 (Markovic.svetomir@mayo.edu).

## REFERENCES

- Krebs ET Jr, Krebs ET Sr, Beard HH. The Unitarian or Trophoblastic Thesis of Cancer. *Med Rec.* 1950;163(7):149-174.
- Beard HH. Correlation of the unitarian or trophoblastic thesis with the biological test of malignancy. *Fed Proc.* 1948;7(1, Pt 1):145.
- Krebs ET Jr. Carcinogenesis; in the light of the trophoblastic or unitarian thesis of cancer. *Int Rec Med Gen Pract Clin.* 1951; 164(3):141-169; contd.
- Ferretti C, Bruni L, Dangles-Marie V, Pecking AP, Bellet D. Molecular circuits shared by placental and cancer cells, and their implications in the proliferative, invasive and migratory capacities of trophoblasts. *Hum Reprod Update.* 2007;13(2): 121-141.
- Holtan SG, Creedon DJ, Haluska P, Markovic SN. Cancer and pregnancy: parallels in growth, invasion, and immune modulation and implications for cancer therapeutic agents. *Mayo Clin Proc.* 2009;84(11):985-1000.
- Park DW, Yang KM. Hormonal regulation of uterine chemokines and immune cells. *Clin Exp Reprod Med.* 2011;38(4):179-185.
- Belfiore A, Perks CM. Grand challenges in cancer endocrinology: endocrine related cancers, an expanding concept. *Front Endocrinol (Lausanne).* 2013;4:141.
- Holtan SG, Mansfield AS, Creedon DJ, et al. An organ system based approach to prognosis in advanced melanoma. *Front Biosci (Elite Ed).* 2012;4:2823-2833.
- Soeters PB, Grimble RF. The conditional role of inflammation in pregnancy and cancer. *Clin Nutr.* 2012;32(3): 460-465.
- Coventry BJ, Ashdown ML, Quinn MA, Markovic SN, Yatomi-Clarke SL, Robinson AP. CRP identifies homeostatic immune oscillations in cancer patients: a potential treatment targeting tool? *J Transl Med.* 2009;7:102.
- Leontovich AA, Dronca RS, Suman VJ, et al. Fluctuation of systemic immunity in melanoma and implications for timing of therapy. *Front Biosci (Elite Ed).* 2012;4:958-975.
- Dronca RS, Leontovich AA, Nevala WK, Markovic SN. Personalized therapy for metastatic melanoma: could timing be everything? *Future Oncol.* 2012;8(11):1401-1406.
- Kirkpatrick HF, Robertson JD. Hormonal changes in pregnancy. *Med Illus.* 1953;7(7):553-555.
- Slominski AT, Zmijewski MA, Skobowiat C, Zbytek B, Slominski RM, Steketee JD. Sensing the environment: regulation of local and global homeostasis by the skin's neuroendocrine system. *Adv Anat Embryol Cell Biol.* 2012;212:v-115.
- Zouboulis CC, Chen WC, Thomson MJ, Qin K, Rosenfield R. Sexual hormones in human skin. *Horm Metab Res.* 2007;39(2): 85-95.
- Slominski A, Zmijewski MA, Pawelek J. L-tyrosine and L-dihydroxyphenylalanine as hormone-like regulators of melanocyte functions. *Pigment Cell Mel Res.* 2012;25(1):14-27.
- Slominski A, Tobin DJ, Shibahara S, Wortsman J. Melanin pigmentation in mammalian skin and its hormonal regulation. *Physiol Rev.* 2004;84(4):1155-1228.
- Osterlind A, Tucker MA, Stone BJ, Jensen OM. The Danish case-control study of cutaneous malignant melanoma, III: hormonal and reproductive factors in women. *Int J Cancer.* 1988; 42(6):821-824.
- Koomen ER, Joosse A, Herings RM, Casparie MK, Guchelaar HJ, Nijsten T. Estrogens, oral contraceptives and hormonal replacement therapy increase the incidence of cutaneous melanoma: a population-based case-control study. *Ann Oncol.* 2009;20(2):358-364.
- Lens M, Bataille V. Melanoma in relation to reproductive and hormonal factors in women: current review on controversial issues. *Cancer Causes Control.* 2008;19(5):437-442.
- Stensheim H, Møller B, van Dijk T, Fosså SD. Cause-specific survival for women diagnosed with cancer during pregnancy or lactation: a registry-based cohort study. *J Clin Oncol.* 2009;27(1):45-51.
- Mansfield AS, Nevala WK, Dronca RS, Leontovich AA, Shuster L, Markovic SN. Normal ageing is associated with an increase in Th2 cells, MCP-1 (CCL1) and RANTES (CCL5), with differences in sCD40L and PDGF-AA between sexes. *Clin Exp Immunol.* 2012;170(2):186-193.
- Beddingfield FC III. The melanoma epidemic: res ipsa loquitur. *Oncologist.* 2003;8(5):459-465.
- Balch CM, Soong SJ, Gershenwald JE, et al. Prognostic factors analysis of 17,600 melanoma patients: validation of the American Joint Committee on Cancer melanoma staging system. *J Clin Oncol.* 2001;19(16):3622-3634.
- Kemeny MM, Busch E, Stewart AK, Menck HR. Superior survival of young women with malignant melanoma. *Am J Surg.* 1998;175(6):437-444; discussion 444-445.
- Shaw HM, Milton GW, Farago G, McCarthy WH. Endocrine influences on survival from malignant melanoma. *Cancer.* 1978;42(2):669-677.
- Mervic L. Time course and pattern of metastasis of cutaneous melanoma differ between men and women. *PLoS One.* 2012; 7(3):e32955.
- Ticconi C, Zicari A, Belmonte A, Realacci M, Rao ChV, Piccione E. Pregnancy-promoting actions of hCG in human myometrium and fetal membranes. *Placenta.* 2007;28(Suppl A):S137-S143.

29. Keay SD, Vatsish M, Karteris E, Hillhouse EW, Randeve HS. The role of hCG in reproductive medicine. *BJOG*. 2004;111(11):1218-1228.
30. Wan H, Versnel MA, Cheung WY, et al. Chorionic gonadotropin can enhance innate immunity by stimulating macrophage function. *J Leukoc Biol*. 2007;82(4):926-933.
31. Schumacher A, Brachwitz N, Sohr S, et al. Human chorionic gonadotropin attracts regulatory T cells into the fetal-maternal interface during early human pregnancy. *J Immunol*. 2009;182(9):5488-5497.
32. Tsampalas M, Grیدهlet V, Berndt S, Foidart JM, Geenen V, Perrier d'Hauterive S. Human chorionic gonadotropin: a hormone with immunological and angiogenic properties. *J Reprod Immunol*. 2010;85(1):93-98.
33. Sherwin JR, Sharkey AM, Cameo P, et al. Identification of novel genes regulated by chorionic gonadotropin in baboon endometrium during the window of implantation. *Endocrinology*. 2007;148(2):618-626.
34. Kane N, Kelly R, Saunders PT, Critchley HO. Proliferation of uterine natural killer cells is induced by human chorionic gonadotropin and mediated via the mannose receptor. *Endocrinology*. 2009;150(6):2882-2888.
35. Moffett A, Loke C. Immunology of placentation in eutherian mammals. *Nat Rev Immunol*. 2006;6(8):584-594.
36. Vacca P, Moretta L, Moretta A, Mingari MC. Origin, phenotype and function of human natural killer cells in pregnancy. *Trends Immunol*. 2011;32(11):517-523.
37. Holtan SG, Creedon DJ, Thompson MA, Nevala WK, Markovic SN. Expansion of CD16-negative natural killer cells in the peripheral blood of patients with metastatic melanoma. *Clin Dev Immunol*. 2011;2011:316314.
38. Koldehoff M, Katzorke T, Wisbrun NC, et al. Modulating impact of human chorionic gonadotropin hormone on the maturation and function of hematopoietic cells. *J Leukoc Biol*. 2011;90(5):1017-1026.
39. Reisinger K, Baal N, McKinnon T, Munstedt K, Zygmunt M. The gonadotropins: tissue-specific angiogenic factors? *Mol Cell Endocrinol*. 2007;269(1-2):65-80.
40. Kalkunte SS, Mselle TF, Norris WE, Wira CR, Sentman CL, Sharma S. Vascular endothelial growth factor C facilitates immune tolerance and endothelial activity of human uterine NK cells at the maternal-fetal interface. *J Immunol*. 2009;182(7):4085-4092.
41. Berndt S, Perrier d'Hauterive S, Blacher S, et al. Angiogenic activity of human chorionic gonadotropin through LH receptor activation on endothelial and epithelial cells of the endometrium. *FASEB J*. 2006;20(14):2630-2632.
42. Thaxton JE, Sharma S. Interleukin-10: a multi-faceted agent of pregnancy. *Am J Reprod Immunol*. 2010;63(6):482-491.
43. Brouillet S, Hoffmann P, Chauvet S, et al. Revisiting the role of hCG: new regulation of the angiogenic factor EG-VEGF and its receptors. *Cell Mol Life Sci*. 2012;69(9):1537-1550.
44. Kayisli UA, Selam B, Guzeloglu-Kayisli O, Demir R, Arici A. Human chorionic gonadotropin contributes to maternal immunotolerance and endometrial apoptosis by regulating Fas-Fas ligand system. *J Immunol*. 2003;171(5):2305-2313.
45. Lei ZM, Rao CV, Ackerman DM, Day TG. The expression of human chorionic gonadotropin/human luteinizing hormone receptors in human gestational trophoblastic neoplasms. *J Clin Endocrinol Metab*. 1992;74(6):1236-1241.
46. Sarandakou A, Protonotariou E, Rizos D. Tumor markers in biological fluids associated with pregnancy. *Crit Rev Clin Lab Sci*. 2007;44(2):151-178.
47. Cole LA. hCG, the wonder of today's science. *Reprod Biol Endocrinol*. 2012;10:24.
48. Doi F, Chi DD, Charuwom BB, et al. Detection of beta-human chorionic gonadotropin mRNA as a marker for cutaneous malignant melanoma. *Int J Cancer*. 1996;65(4):454-459.
49. Ayala AR, Saad A, Vázquez X, Ramirez-Wiella G, Perches RD. Human chorionic gonadotropin immunoreactivity in serum of patients with malignant neoplasms. *Am J Reprod Immunol*. 1983;3(3):149-151.
50. Akhvediani L, Nagervadze M, Dumbadze G. Detection of tolerance against human chorionic gonadotropin at malignant and benign tumors of female reproduction system. *Georgian Med News*. 2009;171:20-24.
51. Butler SA, Ikram MS, Mathieu S, Iles RK. The increase in bladder carcinoma cell population induced by the free beta subunit of human chorionic gonadotropin is a result of an anti-apoptosis effect and not cell proliferation. *Br J Cancer*. 2000;82(9):1553-1556.
52. Zygmunt M, Herr F, Keller-Schoenwetter S, et al. Characterization of human chorionic gonadotropin as a novel angiogenic factor. *J Clin Endocrinol Metab*. 2002;87(11):5290-5296.
53. Lin H, Mosmann TR, Guilbert L, Tuntipopipat S, Wegmann TG. Synthesis of T helper 2-type cytokines at the maternal-fetal interface. *J Immunol*. 1993;151(9):4562-4573.
54. Szekeres-Bartho J, Halasz M, Palkovics T. Progesterone in pregnancy; receptor-ligand interaction and signaling pathways. *J Reprod Immunol*. 2009;83(1-2):60-64.
55. Szekeres-Bartho J, Barakonyi A, Polgar B, et al. The role of gamma/delta T cells in progesterone-mediated immunomodulation during pregnancy: a review. *Am J Reprod Immunol*. 1999;42(1):44-48.
56. Szekeres-Bartho J, Polgar B. PIBF: the double edged sword. Pregnancy and tumor. *Am J Reprod Immunol*. 2010;64(2):77-86.
57. Gadkar-Sable S, Shah C, Rosario G, Sachdeva G, Puri C. Progesterone receptors: various forms and functions in reproductive tissues. *Front Biosci*. 2005;10:2118-2130.
58. Tan H, Yi L, Rote NS, Hurd WW, Mesiano S. Progesterone receptor-A and -B have opposite effects on proinflammatory gene expression in human myometrial cells: implications for progesterone actions in human pregnancy and parturition. *J Clin Endocrinol Metab*. 2012;97(5):E719-E730.
59. Mesiano S, Chan EC, Fitter JT, Kwek K, Yeo G, Smith R. Progesterone withdrawal and estrogen activation in human parturition are coordinated by progesterone receptor A expression in the myometrium. *J Clin Endocrinol Metab*. 2002;87(6):2924-2930.
60. Vrachnis N, Malamas FM, Sifakis S, Tsikouras P, Iliodromiti Z. Immune aspects and myometrial actions of progesterone and CRH in labor. *Clin Dev Immunol*. 2012;2012:937618.
61. Ishida M, Choi JH, Hirabayashi K, et al. Reproductive phenotypes in mice with targeted disruption of the 20alpha-hydroxysteroid dehydrogenase gene. *J Reprod Dev*. 2007;53(3):499-508.
62. Arck PC, Rücke M, Rose M, et al. Early risk factors for miscarriage: a prospective cohort study in pregnant women. *Reprod Biomed Online*. 2008;17(1):101-113.
63. Szekeres-Bartho J, Hadnagy J, Pacsa AS. The suppressive effect of progesterone on lymphocyte cytotoxicity: unique progesterone sensitivity of pregnancy lymphocytes. *J Reprod Immunol*. 1985;7(2):121-128.
64. Liang J, Sun L, Wang Q, Hou Y. Progesterone regulates mouse dendritic cells differentiation and maturation. *Int Immunopharmacol*. 2006;6(5):830-838.
65. De León-Nava MA, Nava K, Soldevila G, et al. Immune sexual dimorphism: effect of gonadal steroids on the expression of cytokines, sex steroid receptors, and lymphocyte proliferation. *J Steroid Biochem Mol Biol*. 2009;113(1-2):57-64.
66. Piccinni MP, Giudizi MG, Biagiotti R, et al. Progesterone favors the development of human T helper cells producing Th2-type cytokines and promotes both IL-4 production and membrane CD30 expression in established Th1 cell clones. *J Immunol*. 1995;155(1):128-133.
67. Canellada A, Blois S, Gentile T, Margni Idehu RA. In vitro modulation of protective antibody responses by estrogen, progesterone and interleukin-6. *Am J Reprod Immunol*. 2002;48(5):334-343.



68. Slominski A, Gomez-Sanchez CE, Foecking MF, Wortsman J. Metabolism of progesterone to DOC, corticosterone and 18OHDOC in cultured human melanoma cells. *FEBS Lett*. 1999;455(3):364-366.
69. Slominski A, Ermak G, Mihm M. ACTH receptor, CYP11A1, CYP17 and CYP21A2 genes are expressed in skin. *J Clin Endocrinol Metab*. 1996;81(7):2746-2749.
70. Slominski A, Zjawiony J, Wortsman J, et al. A novel pathway for sequential transformation of 7-dehydrocholesterol and expression of the P450<sub>scc</sub> system in mammalian skin. *Eur J Biochem*. 2004;271(21):4178-4188.
71. Wiedemann C, Nägele U, Schramm G, Berking C. Inhibitory effects of progestogens on the estrogen stimulation of melanocytes in vitro. *Contraception*. 2009;80(3):292-298.
72. Im S, Lee ES, Kim W, et al. Donor specific response of estrogen and progesterone on cultured human melanocytes. *J Korean Med Sci*. 2002;17(1):58-64.
73. Fang X, Zhang X, Zhou M, Li J. Effects of progesterone on the growth regulation in classical progesterone receptor-negative malignant melanoma cells. *J Huazhong Univ Sci Technolog Med Sci*. 2010;30(2):231-234.
74. Kozma N, Halasz M, Polgar B, et al. Progesterone-induced blocking factor activates STAT6 via binding to a novel IL-4 receptor. *J Immunol*. 2006;176(2):819-826.
75. Kanda N, Watanabe S. 17beta-estradiol, progesterone, and dihydrotestosterone suppress the growth of human melanoma by inhibiting interleukin-8 production. *J Invest Dermatol*. 2001;117(2):274-283.
76. Nevala WK, Vachon CM, Leontovich AA, Scott CG, Thompson MA, Markovic SN, Melanoma Study Group of the Mayo Clinic Cancer Center. Evidence of systemic Th2-driven chronic inflammation in patients with metastatic melanoma. *Clin Cancer Res*. 2009;15(6):1931-1939.
77. Carmeliet P, Moons L, Luttun A, et al. Synergism between vascular endothelial growth factor and placental growth factor contributes to angiogenesis and plasma extravasation in pathological conditions. *Nat Med*. 2001;7(5):575-583.
78. Clauss M, Weich H, Breier G, et al. The vascular endothelial growth factor receptor Flt-1 mediates biological activities. Implications for a functional role of placenta growth factor in monocyte activation and chemotaxis. *J Biol Chem*. 1996;271(30):17629-17634.
79. Selvaraj SK, Giri RK, Perelman N, Johnson C, Malik P, Kalra VK. Mechanism of monocyte activation and expression of proinflammatory cytochemokines by placenta growth factor. *Blood*. 2003;102(4):1515-1524.
80. Bagley RG, Ren Y, Weber W, et al. Placental growth factor upregulation is a host response to antiangiogenic therapy. *Clin Cancer Res*. 2011;17(5):976-988.
81. Fischer C, Jonckx B, Mazzone M, et al. Anti-PlGF inhibits growth of VEGF(R)-inhibitor-resistant tumors without affecting healthy vessels. *Cell*. 2007;131(3):463-475.
82. Eriksson A, Cao R, Pawliuk R, et al. Placenta growth factor-1 antagonizes VEGF-induced angiogenesis and tumor growth by the formation of functionally inactive PlGF-1/VEGF heterodimers. *Cancer Cell*. 2002;1(1):99-108.
83. Loges S, Schmidt T, Carmeliet P. "Antimyoangiogenic" therapy for cancer by inhibiting PlGF. *Clin Cancer Res*. 2009;15(11):3648-3653.
84. Bryant-Greenwood GD, Yamamoto SY, Sadowsky DW, Gravett MG, Novy MJ. Relaxin stimulates interleukin-6 and interleukin-8 secretion from the extraplacental chorionic cytotrophoblast. *Placenta*. 2009;30(7):599-606.
85. Tashima LS, Mazoujian G, Bryant-Greenwood GD. Human relaxin in normal, benign and neoplastic breast tissue. *J Mol Endocrinol*. 1994;12(3):351-364.
86. Mazoujian G, Bryant-Greenwood GD. Relaxin in breast tissue. *Lancet*. 1990;335(8684):298-299.
87. Hansell DJ, Bryant-Greenwood GD, Greenwood FC. Expression of the human relaxin HI gene in the decidua, trophoblast, and prostate. *J Clin Endocrinol Metab*. 1991;72(4):899-904.
88. Perkins AV, Wolfe CD, Eben F, Soothill P, Linton EA. Corticotrophin-releasing hormone-binding protein in human fetal plasma. *J Endocrinol*. 1995;146(3):395-401.
89. Grammatopoulos DK, Hillhouse EW. Role of corticotropin-releasing hormone in onset of labour. *Lancet*. 1999;354(9189):1546-1549.
90. Slominski A, Wortsman J, Luger T, Paus R, Solomon S. Corticotropin releasing hormone and proopiomelanocortin involvement in the cutaneous response to stress. *Physiol Rev*. 2000;80(3):979-1020.
91. Slominski A, Zbytek B, Szczesniowski A, et al. CRH stimulation of corticosteroids production in melanocytes is mediated by ACTH. *Am J Physiol Endocrinol Metab*. 2005;288(4):E701-E706.
92. Yang Y, Park H, Yang Y, Kim TS, Bang SI, Cho D. Enhancement of cell migration by corticotropin-releasing hormone through ERK1/2 pathway in murine melanoma cell line, B16F10. *Exp Dermatol*. 2007;16(1):22-27.
93. Kim MH, Cho D, Kim HJ, et al. Investigation of the corticotropin-releasing hormone-proopiomelanocortin axis in various skin tumours. *Br J Dermatol*. 2006;155(5):910-915.
94. Slominski A, Zbytek B, Zmijewski M, et al. Corticotropin releasing hormone and the skin. *Front Biosci*. 2006;11:2230-2248.
95. Slominski AT, Zmijewski MA, Zbytek B, Tobin DJ, Theoharides TC, Rivier J. Key role of CRF in the skin stress response system. *Endocr Rev*. 2013;34(6):827-884.
96. Carlson KW, Nawy SS, Wei ET, et al. Inhibition of mouse melanoma cell proliferation by corticotropin-releasing hormone and its analogs. *Anticancer Res*. 2001;21(2A):1173-1179.
97. Slominski A, Zbytek B, Pisarchik A, Slominski RM, Zmijewski MA, Wortsman J. CRH functions as a growth factor/cytokine in the skin. *J Cell Physiol*. 2006;206(3):780-791.
98. Chrousos GP. The hypothalamic-pituitary-adrenal axis and immune-mediated inflammation. *N Engl J Med*. 1995;332(20):1351-1362.
99. Ilias I, Mastorakos G. The emerging role of peripheral corticotropin-releasing hormone (CRH). *J Endocrinol Invest*. 2003;26(4):364-371.
100. Ciereszko RE, Petroff BK, Ottobre AC, Guan Z, Stokes BT, Ottobre JS. Assessment of the mechanism by which prolactin stimulates progesterone production by early corpora lutea of pigs. *J Endocrinol*. 1998;159(2):201-209.
101. Herrera E, Muñoz C, López-Luna P, Ramos P. Carbohydrate-lipid interactions during gestation and their control by insulin. *Braz J Med Biol Res*. 1994;27(11):2499-2519.
102. Sorenson RL, Brelje TC. Adaptation of islets of Langerhans to pregnancy: beta-cell growth, enhanced insulin secretion and the role of lactogenic hormones. *Horm Metab Res*. 1997;29(6):301-307.
103. Lellis-Santos C, Sakamoto LH, Bromati CR, et al. The regulation of Rasd1 expression by glucocorticoids and prolactin controls peripartum maternal insulin secretion. *Endocrinology*. 2012;153(8):3668-3678.
104. Voegt JL, Lee Y, Yang S, Arbogast L. Regulation of prolactin secretion during pregnancy and lactation. *Prog Brain Res*. 2001;133:173-185.
105. Martini JF, Piot C, Humeau LM, Struman I, Martial JA, Weiner RI. The antiangiogenic factor I6K PRL induces programmed cell death in endothelial cells by caspase activation. *Mol Endocrinol*. 2000;14(10):1536-1549.
106. Hilfiker-Kleiner D, Kaminski K, Podewski E, et al. A cathepsin D-cleaved 16 kDa form of prolactin mediates postpartum cardiomyopathy. *Cell*. 2007;128(3):589-600.
107. Matera L, Contarini M, Bellone G, Forno B, Biglino A. Up-modulation of interferon-gamma mediates the enhancement of spontaneous cytotoxicity in prolactin-activated natural killer cells. *Immunology*. 1999;98(3):386-392.



108. Peeva E, Zouali M. Spotlight on the role of hormonal factors in the emergence of autoreactive B-lymphocytes. *Immunol Lett*. 2005;101(2):123-143.
109. Chavez-Rueda K, Hernández J, Zenteno E, Leños-Miranda A, Legorreta-Haquet MV, Blanco-Favela F. Identification of prolactin as a novel immunomodulator on the expression of co-stimulatory molecules and cytokine secretions on T and B human lymphocytes. *Clin Immunol*. 2005;116(2):182-191.
110. Yu-Lee LY. Prolactin modulation of immune and inflammatory responses. *Recent Prog Horm Res*. 2002;57:435-455.
111. Orbach H, Zandman-Goddard G, Amital H, et al. Novel biomarkers in autoimmune diseases: prolactin, ferritin, vitamin D, and TPA levels in autoimmune diseases. *Ann N Y Acad Sci*. 2007;1109:385-400.
112. Majumder B, Biswas R, Chattopadhyay U. Prolactin regulates antitumor immune response through induction of tumoricidal macrophages and release of IL-12. *Int J Cancer*. 2002;97(4):493-500.
113. Sun R, Wei H, Zhang J, Li A, Zhang W, Tian Z. Recombinant human prolactin improves antitumor effects of murine natural killer cells in vitro and in vivo. *Neuroimmunomodulation*. 2002-2003;10(3):169-176.
114. Swaminathan G, Varghese B, Fuchs SY. Regulation of prolactin receptor levels and activity in breast cancer. *J Mammary Gland Biol Neoplasia*. 2008;13(1):81-91.
115. Asher JM, O'Leary KA, Rugowski DE, Arendt LM, Schuler LA. Prolactin promotes mammary pathogenesis independently from cyclin d1. *Am J Pathol*. 2012;181(1):294-302.
116. Chikanza IC. Prolactin and neuroimmunomodulation: in vitro and in vivo observations. *Ann N Y Acad Sci*. 1999;876:119-130.
117. Bole-Feyso C, Goffin V, Edery M, Binart N, Kelly PA. Prolactin (PRL) and its receptor: actions, signal transduction pathways and phenotypes observed in PRL receptor knockout mice. *Endocr Rev*. 1998;19(3):225-268.
118. Peverelli E, Mantovani G, Vitali E, et al. Filamin-A is essential for dopamine D2 receptor expression and signaling in tumorous lactotrophs. *J Clin Endocrinol Metab*. 2012;97(3):967-977.
119. Ben-Jonathan N, Hnasko R. Dopamine as a prolactin (PRL) inhibitor. *Endocr Rev*. 2001;22(6):724-763.
120. Richards SM, Murphy WJ. Use of human prolactin as a therapeutic protein to potentiate immunohematopoietic function. *J Neuroimmunol*. 2000;109(1):56-62.
121. Tabruyn SP, Sabatel C, Nguyen NQ, et al. The angiostatic I6K human prolactin overcomes endothelial cell anergy and promotes leukocyte infiltration via nuclear factor-kappaB activation. *Mol Endocrinol*. 2007;21(6):1422-1429.
122. Nguyen NQ, Castermans K, Berndt S, et al. The antiangiogenic I6K prolactin impairs functional tumor neovascularization by inhibiting vessel maturation. *PLoS One*. 2011;6(11):e27318.
123. Cameliet P, Jain RK. Angiogenesis in cancer and other diseases. *Nature*. 2000;407(6801):249-257.
124. Samal B, Sun Y, Stearns G, Xie C, Suggs S, McNiece I. Cloning and characterization of the cDNA encoding a novel human pre-B-cell colony-enhancing factor. *Mol Cell Biol*. 1994;14(2):1431-1437.
125. Rongvaux A, Shea RJ, Mulks MH, et al. Pre-B-cell colony-enhancing factor, whose expression is up-regulated in activated lymphocytes, is a nicotinamide phosphoribosyltransferase, a cytosolic enzyme involved in NAD biosynthesis. *Eur J Immunol*. 2002;32(11):3225-3234.
126. Moschen AR, Kaser A, Enrich B, et al. Visfatin, an adipocytokine with proinflammatory and immunomodulating properties. *J Immunol*. 2007;178(3):1748-1758.
127. Ognjanovic S, Tashima LS, Bryant-Greenwood GD. The effects of pre-B-cell colony-enhancing factor on the human fetal membranes by microarray analysis. *Am J Obstet Gynecol*. 2003;189(4):1187-1195.
128. Lappas M. Visfatin regulates the terminal processes of human labour and delivery via activation of the nuclear factor-kappaB pathway. *Mol Cell Endocrinol*. 2012;348(1):128-134.
129. Mazaki-Tovi S, Romero R, Vaisbuch E, et al. Maternal plasma visfatin in preterm labor. *J Matern Fetal Neonatal Med*. 2009;22(8):693-704.
130. Maldí E, Travelli C, Caldarelli A, et al. Nicotinamide phosphoribosyltransferase (NAMPT) is over-expressed in melanoma lesions. *Pigment Cell Melanoma Res*. 2013;26(1):144-146.
131. Buldak RJ, Buldak Ł, Polaniak R, et al. Visfatin affects redox adaptive responses and proliferation in Me45 human malignant melanoma cells: an in vitro study. *Oncol Rep*. 2013;29(2):771-778.
132. Jia SH, Li Y, Parodo J, et al. Pre-B cell colony-enhancing factor inhibits neutrophil apoptosis in experimental inflammation and clinical sepsis. *J Clin Invest*. 2004;113(9):1318-1327.
133. Tanriverdi F, Silveira LF, MacColl GS, Bouloux PM. The hypothalamic-pituitary-gonadal axis: immune function and autoimmunity. *J Endocrinol*. 2003;176(3):293-304.
134. Cohen-Solal JF, Jegathanan V, Grimaldi CM, Peeva E, Diamond B. Sex hormones and SLE: influencing the fate of autoreactive B cells. *Curr Top Microbiol Immunol*. 2006;305:67-88.
135. Straub RH. The complex role of estrogens in inflammation. *Endocr Rev*. 2007;28(5):521-574.
136. Zang YC, Halder JB, Hong J, Rivera VM, Zhang JZ. Regulatory effects of estradiol on T cell migration and cytokine profile: inhibition of transcription factor NF-kappa B. *J Neuroimmunol*. 2002;124(1-2):106-114.
137. Phiel KL, Henderson RA, Adelman SJ, Elloso MM. Differential estrogen receptor gene expression in human peripheral blood mononuclear cell populations. *Immunol Lett*. 2005;97(1):107-113.
138. Holtan SG, Creedon DJ. Mother knows best: lessons from fetomaternal tolerance applied to cancer immunity. *Front Biosci (Schol Ed)*. 2011;3:1533-1540.
139. Lee YL, Cheong AW, Chow WN, Lee KF, Yeung WS. Regulation of complement-3 protein expression in human and mouse oviducts. *Mol Reprod Dev*. 2009;76(3):301-308.
140. Shifren JL, Tseng JF, Zaloudek CJ, et al. Ovarian steroid regulation of vascular endothelial growth factor in the human endometrium: implications for angiogenesis during the menstrual cycle and in the pathogenesis of endometriosis. *J Clin Endocrinol Metab*. 1996;81(8):3112-3118.
141. Hildebrandt VA, Babischkin JS, Koos RD, Pepe GJ, Albrecht ED. Developmental regulation of vascular endothelial growth/permeability factor messenger ribonucleic acid levels in and vascularization of the villous placenta during baboon pregnancy. *Endocrinology*. 2001;142(5):2050-2057.
142. Schoenberg BS, Christine BW. Malignant melanoma associated with breast cancer. *South Med J*. 1980;73(11):1493-1497.
143. Walker MJ, Beattie CW, Patel MK, Ronan SM, Das Gupta TK. Estrogen receptor in malignant melanoma. *J Clin Oncol*. 1987;5(8):1256-1261.
144. Zen M, Ghirardello A, Iaccarino L, et al. Hormones, immune response, and pregnancy in healthy women and SLE patients. *Swiss Med Wkly*. 2010;140(13-14):187-201.
145. Grimaldi CM, Cleary J, Dagtas AS, Moussai D, Diamond B. Estrogen alters thresholds for B cell apoptosis and activation. *J Clin Invest*. 2002;109(12):1625-1633.
146. Cutolo M, Sulli A, Craviotto C, et al. Modulation of cell growth and apoptosis by sex hormones in cultured monocytic THP-1 cells. *Ann N Y Acad Sci*. 2002;966:204-210.
147. Hughes GC, Clark EA. Regulation of dendritic cells by female sex steroids: relevance to immunity and autoimmunity. *Autoimmunity*. 2007;40(6):470-481.
148. Calippe B, Douin-Echinard V, Laffargue M, et al. Chronic estradiol administration in vivo promotes the proinflammatory response of macrophages to TLR4 activation: involvement of the phosphatidylinositol 3-kinase pathway. *J Immunol*. 2008;180(12):7980-7988.

149. Beagley KW, Gockel CM. Regulation of innate and adaptive immunity by the female sex hormones oestradiol and progesterone. *FEMS Immunol Med Microbiol*. 2003;38(1):13-22.
150. Evans MJ, MacLaughlin S, Marvin RD, Abdou NI. Estrogen decreases in vitro apoptosis of peripheral blood mononuclear cells from women with normal menstrual cycles and decreases TNF-alpha production in SLE but not in normal cultures. *Clin Immunol Immunopathol*. 1997;82(3):258-262.
151. Kuiper GG, Carlsson B, Grandien K, et al. Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. *Endocrinology*. 1997;138(3):863-870.
152. Ohata C, Tadokoro T, Itami S. Expression of estrogen receptor beta in normal skin, melanocytic nevi and malignant melanomas. *J Dermatol*. 2008;35(4):215-221.
153. Schmidt AN, Nanney LB, Boyd AS, King LE Jr, Ellis DL. Oestrogen receptor-beta expression in melanocytic lesions. *Exp Dermatol*. 2006;15(12):971-980.
154. Scolyer RA, Judge MJ, Evans A, et al. Data set for pathology reporting of cutaneous invasive melanoma: recommendations from the International Collaboration on Cancer Reporting (ICCR). *Am J Surg Pathol*. 2013;37(12):1797-1814.
155. de Giorgi V, Mavilia C, Massi D, et al. Estrogen receptor expression in cutaneous melanoma: a real-time reverse transcriptase-polymerase chain reaction and immunohistochemical study. *Arch Dermatol*. 2009;145(1):30-36.
156. Mor G, Sapi E, Abrahams VM, et al. Interaction of the estrogen receptors with the Fas ligand promoter in human monocytes. *J Immunol*. 2003;170(1):114-122.
157. Paharkova-Vatchkova V, Maldonado R, Kovats S. Estrogen preferentially promotes the differentiation of CD11c+ CD11b(intermediate) dendritic cells from bone marrow precursors. *J Immunol*. 2004;172(3):1426-1436.
158. Mao A, Paharkova-Vatchkova V, Hardy J, Miller MM, Kovats S. Estrogen selectively promotes the differentiation of dendritic cells with characteristics of Langerhans cells. *J Immunol*. 2005;175(8):5146-5151.
159. Majeesh NJ, Escuin D, LaVallee TM, et al. ZME2 inhibits tumor growth and angiogenesis by disrupting microtubules and dysregulating HIF. *Cancer Cell*. 2003;3(4):363-375.
160. Miller VM. Sex-based differences in vascular function. *Women's Health (Lond Engl)*. 2010;6(5):737-752.
161. Raaz-Schrauder D, Klinghammer L, Baum C, et al. Association of systemic inflammation markers with the presence and extent of coronary artery calcification. *Cytokine*. 2012;57(2):251-257.
162. Hu FB, Stampfer MJ, Manson JE, et al. Trends in the incidence of coronary heart disease and changes in diet and lifestyle in women. *N Engl J Med*. 2000;343(8):530-537.
163. Pennell LM, Galligan CL, Fish EN. Sex affects immunity. *J Autoimmun*. 2012;38(2-3):282-291.
164. Whitacre CC. Sex differences in autoimmune disease. *Nat Immunol*. 2001;2(9):777-780.
165. Ostensen M, Villiger PM. Immunology of pregnancy—pregnancy as a remission inducing agent in rheumatoid arthritis. *Transpl Immunol*. 2002;9(2-4):155-160.
166. Scotland RS, Stables MJ, Madalli S, Watson P, Gilroy DW. Sex differences in resident immune cell phenotype underlie more efficient acute inflammatory responses in female mice. *Blood*. 2011;118(22):5918-5927.
167. Marriott I, Huet-Hudson YM. Sexual dimorphism in innate immune responses to infectious organisms. *Immunol Res*. 2006;34(3):177-192.
168. Adrie C, Azoulay E, Francois A, et al. Influence of gender on the outcome of severe sepsis: a reappraisal. *Chest*. 2007;132(6):1786-1793.
169. Hill JA, Polgar K, Anderson DJ. T-helper 1-type immunity to trophoblast in women with recurrent spontaneous abortion. *JAMA*. 1995;273(24):1933-1936.
170. Moraru M, Carbone J, Alecsandru D, et al. Intravenous immunoglobulin treatment increased live birth rate in a Spanish cohort of women with recurrent reproductive failure and expanded CD56(+) cells. *Am J Reprod Immunol*. 2012;68(1):75-84.
171. Jin LP, Zhou YH, Wang MY, Zhu XY, Li DJ. Blockade of CD80 and CD86 at the time of implantation inhibits maternal rejection to the allogeneic fetus in abortion-prone matings. *J Reprod Immunol*. 2005;65(2):133-146.
172. Fan W, Li SW, Liu XR. Influence of blockade of costimulation on Th1/Th2 cytokines shift in unexplained early recurrent spontaneous [In Chinese]. *Sichuan Da Xue Xue Bao Yi Xue Ban*. 2006;37(5):773-775.
173. Li W, Li B, Fan W, et al. CTLA4lg gene transfer alleviates abortion in mice by expanding CD4+CD25+ regulatory T cells and inducing indoleamine 2,3-dioxygenase. *J Reprod Immunol*. 2009;80(1-2):1-11.
174. Weber J. Overcoming immunologic tolerance to melanoma: targeting CTLA-4 with ipilimumab (MDX-010). *Oncologist*. 2008;13(Suppl 4):16-25.
175. Hodi FS, O'Day SJ, McDermott DF, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med*. 2010;363(8):711-723.
176. Trinh VA, Hagen B. Ipilimumab for advanced melanoma: a pharmacologic perspective. *J Oncol Pharm Pract*. 2013;19(3):195-201.
177. Dulos J, Carven GJ, van Bostel SJ, et al. PD-1 blockade augments Th1 and Th17 and suppresses Th2 responses in peripheral blood from patients with prostate and advanced melanoma cancer. *J Immunother*. 2012;35(2):169-178.
178. Hamid O, Robert C, Daud A, et al. Safety and tumor responses with lambrolizumab (anti-PD-1) in melanoma. *N Engl J Med*. 2013;369(2):134-144.
179. Wolchok JD, Kluger H, Callahan MK, et al. Nivolumab plus ipilimumab in advanced melanoma. *N Engl J Med*. 2013;369(2):122-133.
180. Sugiyama D, Nishikawa H, Maeda Y, et al. Anti-CCR4 mAb selectively depletes effector-type FoxP3+CD4+ regulatory T cells, evoking antitumor immune responses in humans. *Proc Natl Acad Sci U S A*. 2013;110(44):17945-17950.
181. Ma J, Han H, Liu D, et al. HER2 as a promising target for cytotoxicity T cells in human melanoma therapy. *PLoS One*. 2013;8(8):e73261.
182. Mansfield AS, Nevala WK, Lieser EA, Leontovich AA, Markovic SN. The immunomodulatory effects of bevacizumab on systemic immunity in patients with metastatic melanoma. *Oncoimmunology*. 2013;2(5):e24436.
183. Gajewski TF, Meng Y, Blank C, et al. Immune resistance orchestrated by the tumor microenvironment. *Immunol Rev*. 2006;213:131-145.
184. Slominski A, Zbytek B, Slominski R. Inhibitors of melanogenesis increase toxicity of cyclophosphamide and lymphocytes against melanoma cells. *Int J Cancer*. 2009;124(6):1470-1477.
185. Slominski A, Zbytek B, Nikolakis G, et al. Steroidogenesis in the skin: implications for local immune functions. *J Steroid Biochem Mol Biol*. 2013;137:107-123.
186. Mordoh J, Tapia IJ, Barrio MM. A word of caution: do not wake sleeping dogs; micrometastases of melanoma suddenly grew after progesterone treatment. *BMC Cancer*. 2013;13:132.
187. Curiel TJ, Wei S, Dong H, et al. Blockade of B7-H1 improves myeloid dendritic cell-mediated antitumor immunity. *Nat Med*. 2003;9(5):562-567.
188. Wang L, Pino-Lagos K, de Vries VC, Guleria I, Sayegh MH, Noelle RJ. Programmed death 1 ligand signaling regulates the generation of adaptive Foxp3+CD4+ regulatory T cells. *Proc Natl Acad Sci U S A*. 2008;105(27):9331-9336.
189. Lin PY, Sun L, Thibodeaux SR, et al. B7-H1-dependent sex-related differences in tumor immunity and immunotherapy responses. *J Immunol*. 2010;185(5):2747-2753.
190. Topalian SL, Hodi FS, Brahmer JR, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med*. 2012;366(26):2443-2454.